

PREVALENCE AND ANTIBIOTIC SENSITIVITY PROFILE OF *Salmonella* SPECIES IN EGGS FROM POULTRY FARMS IN UMUDIKE, ABIA STATE

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ABSTRACT

The increasing reports of the role of poultry products and eggs in the transmission of food borne salmonellosis around the world are raising international public health concern. In this regard a survey was carried out to determine the prevalence and antibiotic profile of Salmonella species in eggs from poultry farms around Umudike, Abia State. A total of two hundred and forty (240) eggs were sampled from four poultry farms. Salmonella species were isolated from 58 of the 240 eggs. The prevalence rates ranged between 8 and 45% with an overall prevalence of 24% of salmonella contamination of the eggs. The antibiotic sensitivity testing was carried out using the disc agar diffusion technique. All the 58 isolates were sensitive to Nitrofurantoin, 42 (72.4%) were sensitive to Gentamicin, 22 (37.9%) to Ofloxacin, 17 (29.3%) to Nalidixic acid while only 4 (6.9%) were sensitive to both Amoxicillin and Tetracycline. All isolates were resistant to Cotrimoxazole. The level of contamination of eggs with Salmonella species in this study calls for urgent need to control the level of Salmonella contamination of poultry farms in the study area. The high level of resistance of the isolates to commonly used antibiotics is really alarming and has great public health significance.

Key words: Antibiotic sensitivity profile, Salmonellosis, Eggs, *Salmonella* spp

INTRODUCTION

Infection of humans with non-typhoid salmonellae from animal sources is an emerging public health problem of significant economic and health burdens in both developed and developing countries (Doughari, *et al.* 2007). An estimated 1.4 millions cases of Salmonellosis involving 16,000 hospitalizations and nearly 600 deaths occurred each year in the United States (Zhao *et al.*, 2003). Estimates of the total patient-related cost of salmonellosis in 1998 in the United States of America ranged from \$275 million to \$1.1 billion (Irwin *et al.*, 1998).

Poultry, poultry products and eggs are major sources of Salmonella-related foodborne illness (Boonmar *et al.*, 1998, Pignato *et al.*, 1995, Irwin *et al.*, 1998, Stevens *et al.*, 1989). International concern over rising isolations of Salmonellae, and

in particular, *Salmonella enteritidis* among humans, and the association with poultry and poultry products has led to the call for establishment of measures to control *Salmonella* in the poultry industry (Rodrigue *et al.*, 1990). Programmes for active surveys of prevalence of *Salmonella* in poultry and poultry products were active in some developed countries such as Canada, (Irwin *et al.*, 1998), Italy (Nastasi *et al.*, 1997, Germany (Schroeter *et al.*, 1994), USA (USDA, 1994), Britain (Humphrey *et al.*, 1988) and more advanced developing countries like Thailand (Sasipreeyajan *et al.*, 1996, Boonmar *et al.*, 2003). There is paucity of data on the prevalence and antimicrobial resistance profile of salmonellae from food animals in the developing countries especially in Sub-Saharan Africa (Okoli *et al.*, 2002). This paper reports a survey

conducted to assess the prevalence and antimicrobial susceptibility profile of *Salmonella* species from eggs from four poultry farms in Umudike, Abia State. Multidrug resistant *Salmonella* serotypes, resistant to commonly used antimicrobial agents have been reported (Zhao *et al.*, 2003, Poppe *et al.*, 1995). Indiscriminate use of antimicrobial agents in clinical and veterinary practices is fuelling the increasing problem of antimicrobial resistance among serotypes of salmonellae. The common practice of using antimicrobial agents in animal production as growth promoters has potential for creating drug resistant strains of bacteria which can be transferred to humans who handle the birds and through the food chain (Okoli *et al.*, 2002). Resistant bacteria can also transfer their resistant genes to other bacteria.

MATERIALS AND METHODS

Sources and Collection of Eggs

A total of 240 eggs were collected from four poultry farms located around Michael Okpara University of Agriculture, Umudike. Sixty (60) eggs were collected, ten eggs on six different days from each of the farms. The eggs were collected into clean plastic bags. The eggs were processed for microbiological analysis at Microbiology Laboratory, Department of Microbiology, Michael Okpara University of Agriculture, Umudike.

Preparation of Culture Media

The culture media used were *Salmonella* – *Shigella* Agar (SSA) (Fluka Biochemika, Fluka Chemie GmbH, India), Muller Hinton Agar (Fluka Biochemika, Fluka Chemie GmbH, Spain) and Selenite F broth Base (Biotec Laboratories Ltd, Martleham Heath Ipswich, UK). All the culture media were prepared according to the manufacturers' instructions.

Isolation of Organisms

The eggs were processed in batches of 10. The shell of each egg was washed with 70% alcohol and then rinsed in sterile

distilled water in a beaker. The egg was transferred to a dry sterile beaker and aseptically broken with sterile forceps. The shell was removed and the content was homogenized using a sterile glass rod. 1ml of the homogenate was introduced into 9ml of Selenite F broth in a sterile culture bottle. Serial dilutions to 10^{-3} were made from this by pipetting 1ml into 9ml of Selenite F broth. The dilutions were incubated overnight. 1ml of the broth culture was then plated out on *Salmonella*-*Shigella* Agar. The SSA plates were read after an overnight incubation at 37°C. The number of colonies on each plate was counted.

Characterization of Isolates

Colonies that showed characteristics of *Salmonella* species on SSA were presumptively identified as *Salmonella* species on the basis of Gram Stain reaction and motility test. Gram Staining and Motility test were done according to standard procedures (Cheesborough, 2000).

Antibiotic Sensitivity Testing

Disc diffusion susceptibility test by Bauer method was carried out as previously described by Ekundayo and Omodamiro (2008). Abtek Sensitivity discs (Abtek Biologicals Ltd, UK) were used.

RESULTS

A total of 240 eggs were collected from four poultry farms in Umudike, Abia State. *Salmonella* species were isolated from 58 eggs. This represents a prevalence of 24.17%. The prevalence of *Salmonella* isolates in the eggs from the farms ranged from 8% to 45%. Table 1 presents the prevalence of *Salmonella* isolates in the eggs from the four farms.

All the fifty-eight isolates were sensitive to Nitrofurantoin and forty-two (72.4%) to Gentamycin. Twenty-two and seventeen of the isolates were sensitive to Ofloxacin and Nalidixic acid respectively. The isolates manifested high level of resistance

to Amoxicillin and Tetracycline with 93.1% of the isolates resistant to both antimicrobial agents. All the isolates were resistant to cotrimoxazole.

Many of the isolates manifested high level of multidrug resistance, sixteen (27.6%) of the isolates were resistant to seven of the antimicrobial agents tested. Thirty-four

(58.6%) were resistant to six of the antimicrobial agents, 44(75.9%) to 5 and 93.1% to 3 antimicrobial agents. The antibiotic sensitivity profile of the 58 Isolates is presented in Table 2. Fig. 3 presents the comparative sensitivity of the isolates to the antimicrobial agents tested.

Table 1: Prevalence of Salmonella isolates in eggs from poultry farms in Umudike

<i>Farm</i>	<i>No. of eggs tested</i>	<i>No. of eggs with growth</i>	% of eggs showing growth
A	60	14	23
B	60	27	45
C	60	5	8
D	60	12	20

Table 2: Antimicrobial Sensitivity profile of Salmonella isolates in eggs from poultry farms in Umudike

Antimicrobial Agents	Disc Potency (μg)	No. (%) of Isolates sensitive	No. (%) of isolates with intermediate sensitivity	No. (%) of isolates Resistant
Amoxycillin	25	4 (6.9)	0 (0)	54 (93.1)
Cotrimoxazole	25	0 (0)	0 (0)	58 (100)
Nitrofurantoin	300	58 (100)	0 (0)	0 (0)
Gentamycin	10	42 (72.4)	0 (0)	16 (27.5)
Ofloxacin	30	22 (37.9)	2 (3.4)	34 (58.6)
Augmentin	30	12 (20.6)	2(3.4)	44 (75.9)
Nalidixic acid	30	17 (29.3)	7 (12.0)	34 (58.6)
Tetracycline	30	4 (6.9)	0 (0)	54 (93.1)

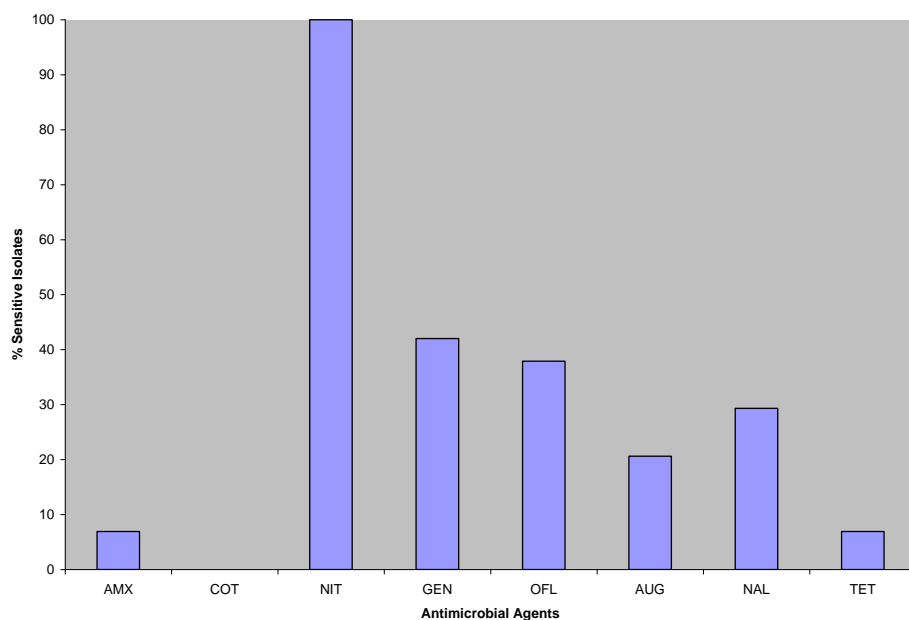


Fig. 1: Percentages of Isolates Sensitive to different Antimicrobial Agents

DISCUSSION

An overall prevalence of 24.17% of *Salmonella* species were isolated from the eggs collected from four farms in Umudike, Abia State. This shows that there is a significant level of contamination of eggs in the four farms from which the eggs were obtained. In one of the farms, up to 45% of the eggs were contaminated with *Salmonella* species. This study only assessed the internal contamination of the eggs. The prevalence of contamination would probably be higher if one takes into consideration the external contamination. The public health impact of contamination of eggs with *Salmonella* species cannot be directly determined from this study. However, reports from advanced countries where public health systems are better organized suggest that transmission of *Salmonella* related food borne illness from eggs is a real public health hazard and that it is associated with huge economic losses and considerable health burdens (Mead *et al.*, 1999, Rodrigue *et al.*, 1990, Stevens *et al.*, 1989, St. Louis *et al.*, 1988). If poultry and egg related *Salmonella* infections remain serious public health problems in developed countries where better sanitation measures and infection controls are in place, it is most likely that the problems are more serious in developing countries. However, there are no data to show the extent of the problem. Most cases of Salmonellosis are self-limiting and in developing countries such diseases are not usually reported, so it is difficult to assess the real impact on the populace. The poultry industry is at infancy in Nigeria, if the expectations of sustainable poultry industry are to be realized, control measures need to be established to control *Salmonella* infections.

The level of antimicrobial resistance of the isolates in this study is really alarming. Many of the isolates are resistant to multiple antimicrobial agents tested. All the isolates were resistant to Cotrimoxazole and 93.1% to Amoxicillin

and Tetracycline. These drugs are the most commonly used antimicrobial agents for treatment of *Salmonella* infections. In a retrospective study covering data for *Salmonella* isolates over a period of four years, Doughari *et al.* (2007) reported a resistance of 88.8% to Cotrimoxazole and various levels of resistance to other drugs in Yola, Northern Nigeria. As in that study, we also recorded high level of resistance to Amoxicillin, 93.1% in this study. Indiscriminate use of antimicrobial agents in medical and veterinary practices is fuelling the antimicrobial resistance problems. The ultimate source of contamination of poultry and animal houses is the environment. Amaechi and Ezeronye (2006) reported high level of contamination of piggery environment with *Salmonella* species in the study area.

CONCLUSION:

This study shows that a considerable level of contamination of eggs occurs in the study area. This study therefore provides the basis for further study on this subject and also the need to assess the public health impact of *Salmonella*-related food borne illness in the poultry industry in Nigeria. There is need to adopt a more efficient hygienic and infection control measures in the poultry industry to control *Salmonella* infections of poultry and prevent the possible public health hazards.

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